

Writing/Speaking Support for 20.109

Atissa Banuazizi

Neal Lerner

Linda Sutliff

Getting to know you: Two truths and one lie

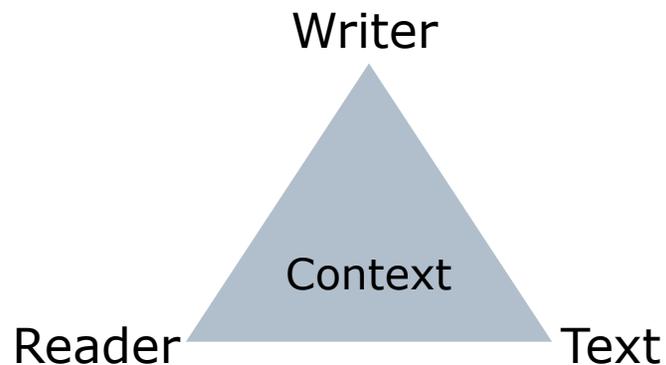
- Write three statements about yourself, two of them true and one a lie.

Getting to know you, part 2: Letter to Atissa, Neal, and Linda

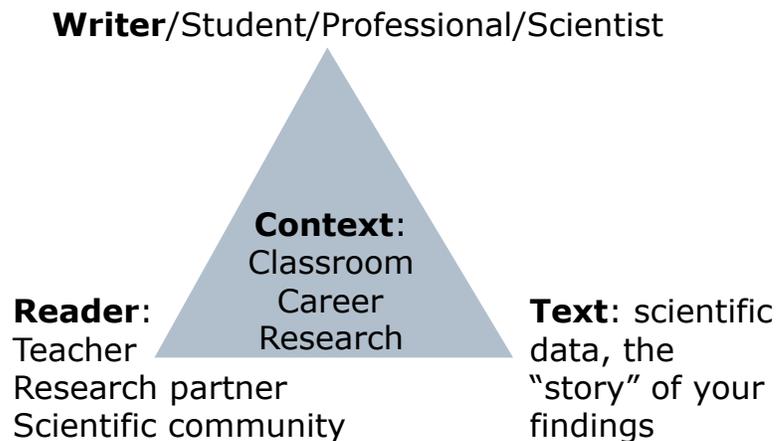
- ❑ What have been your experiences with scientific writing (e.g., lab reports, research articles, reviews)? With other writing tasks?
- ❑ What have been your experiences with giving oral presentations (whether scientific or not)?
- ❑ How would you describe yourself as a writer? As a public speaker?
- ❑ What are your writing/speaking goals for 20.109?

Writing and Thinking Rhetorically about Science

Any writing act can be described in terms of a rhetorical triangle or set of relationships.



The rhetorical relationships for scientific writing can be complex and shifting.



Scientific writers need to control the rhetoric of scientific writing.

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["Empty Rhetoric"](#) by Mike Konopacki

According to Aristotle, rhetoric is "the art of finding in any given case the available means of persuasion."

The goal of scientific writing is to court your audience.

Michael Halloran on Watson & Crick's 1953 "The Structure for DNA"

"The April 1953 paper, then, is really just the initial move in a rhetorical strategy aimed at gaining and holding the attention of an audience. As such, it presumes an understanding of *science as a human community* in which neither facts nor ideas speak for themselves, and the attention of the audience must be courted."

Photo removed due to copyright restrictions.
Watson and Crick with their DNA double helix model.
<http://db2.photoresearchers.com/search/2W6323>

Research article scramble



Photo courtesy of
[Stephanie Booth](#)
on Flickr.

- For the passages from a student's 20.109 laboratory report on homologous recombination: Which section (Introduction, Methods, Results, Discussion, Figure Captions) does each passage belong to?

1.0 Introduction

By obtaining a more profound understanding of all aspects of DNA repair pathways, it may be easier for future breakthroughs in creating chemotherapeutic strategies that specifically and effectively attack cancers, and thus radically change modern cancer treatment. In order to contribute to this understanding of homologous recombination, we have created an assay that will enable us to determine when homologous recombination has taken place.

What features of this paragraph identify it as belong to the Introduction?

The introduction provides a framework for the story you are about to tell, and thus serves two main purposes. For one, you must provide sufficient background information for a reader to understand the forthcoming results. Just as importantly, you must motivate the audience to keep reading! How? Reveal the significance of the work through connections to both prior scientific accomplishments and future applications.

From http://openwetware.org/wiki/20.109%28S10%29:Guidelines_for_writing_up_your_research

The Introduction establishes *context*, *focus*, and *justification*.

Context: Orient your reader to the published literature related to the topic and to essential background information

Focus: Define the research space, stake out territory. What questions are you addressing? What is your hypothesis?

Justification: Show how your work fits into and extends previous work. Argue for the importance of your work.

Swales (1990)

2.0 Methods

In order to perform bacterial transformation, 5 µl of each purification ligation reaction was added to 50 µl of competent bacterial cells, also a positive control was prepared with an uncut pCX-EGFP plasmid. These solutions were then heat shocked in a 42°C bath for 90 seconds so that the competent cells could uptake the DNA. 0.5 ml of LB media was then added to each reaction, and 200 µl of each tube was plated onto separate LB + AMP plates using a sterile spreader. Each plate was then incubated at 37°C overnight.

What features of this paragraph identify it as belong to the Materials & Methods?

The methods section should allow an independent investigator to repeat any of your experiments. Use sub-section headings to allow readers to quickly identify experiments of interest to them (e.g., "Protein conjugation to hydrogels" and "Cell culture and fluorescent labeling"). When commercially available kits were used, it is sufficient to cite the name of the kit and say that it was used according to the manufacturer's protocol. The key to a good methods section is developing your judgment for what information is essential and what is extraneous. Note that the methods section should be written in the past tense, since your experiments are completed at the time you are writing your paper. This section should also be written in complete sentences and paragraphs, not in bullet point form.

From http://openwetware.org/wiki/20.109%28S10%29:Guidelines_for_writing_up_your_research

Your research article should contain a Methods Section, not a Protocol.

A Protocol is . . .

- A series of steps to be carried out.
- Written in sequential or temporal order.
- Intended for the reader to achieve a final result.

A Methods Section is

- A series of steps already completed and is written in past tense.
- Written in logical order.
- Intended for the reader to replicate the experiment.

3.0 Results

As expected the digestion of plasmid backbone (Lane 2) displayed a band of about 4.8 kbp in length, as digesting with SalI would linearize the DNA. However, two other bands were seen in addition to the expected band, which could be due to poor enzyme efficiency. Lanes 3-5 in Figure 6 also confirm the projected length fragments of 3.7 kbp and 1.6 kbp (from Figure 5). This result indicates that the candidate clones were indeed the desired construct.

What features of this paragraph identify it as belong to the Results?

The purpose of the results section is to present your data in a relatively unbiased way, but with some guiding framework. Begin with a short description of the goal and strategy of your overall experiment, and then delve into specific sub-sections that describe each piece of the work.

To write the results section, use the figures and tables as a guide. . . . Present the data as fully as possible, including stuff that does not quite make sense at first glance. Ultimately, each sub-section should begin with an overview sentence that introduces the present experiment and end with a sentence stating the primary conclusion reached from that experiment. (Sub-section headings and figure caption titles can also emphasize said conclusion.) The overview and/or concluding sentences should also provide a transition to the previous/next piece of data when possible. . . .

Note that verbs in the results section are usually in the past tense. Only established scientific knowledge is written about in the present tense, "the world is round," for example. You cannot presume that your own data are part of the body of established scientific knowledge, and so when you describe your own results, use the past tense, "a band of 1.3 kb was seen," for example. There are, however, exceptions to this general rule. It is acceptable to say, "Table 3 shows the sizes of the DNA fragments in our preparation." It is also acceptable to say, "In a 1991 paper, Ebright and coworkers used PCR to mutagenize DNA."

From http://openwetware.org/wiki/20.109%28S10%29:Guidelines_for_writing_up_your_research

What Differentiates Results from the Methods?

Methods = *How*
the data were
accumulated.

Results = *What*
data were
accumulated.

Readers expect to find the "answers" to your research questions in your Results section.

What Differentiates Results from Discussion?

Results = *Data Presentation*

("Experiments showed that")

Discussion = *Data Interpretation*

("Experiments suggest that")

However, you still need to choose which data to present in your Results Section (an act of interpretation!).

4.0 Figure Caption

Results of gel electrophoresis on 1% agarose gel. Lane 1-4 contain the pCX-NNX backbone. In Lane 1 the vector is uncut. In Lane 2 the plasmid is cut with XbaI (? 4.8 kbp), while in Lane 3 it is cut with EcoRI (? 4.8 kbp). Lane 4 shows the backbone double digest with XbaI and EcoRI (?4.7kbp). Lane 5 is the 10Kb DNA Ladder. Lanes 6-7 contain the ?5-EGFP (PCR Product) insert. Lane 6 is the double digest (?0.66 kp), and Lane 7 shows the uncut insert. Lane 8 is the negative PCR-no template control. (Yellow Group W/F)

What features of this paragraph identify it as belong to a Figure caption?

Legends to the figures and tables explain the elements that appear in the illustration. Conclusions about the data are NOT included in the legends. As you write your first draft, you might state in a short simple sentence what the point of the figure or table is. In later drafts, make sure each element of the figure or table is explained. Your figure legends should be written in the present tense since you are explaining elements that still exist at the time that you are writing the paper.

From http://openwetware.org/wiki/20.109%28S10%29:Guidelines_for_writing_up_your_research

Titles and captions allow figures and tables to stand on their own.

- **Guide** the reader to what is most important in the figure.
- **Contextualize** the data shown in terms of purpose and method.
- **Focus attention** on certain findings (e.g., relationship between values).
- **Summarize** the larger point.



Image: public domain
(NASA: Earth as seen from Mars)

Bonus tip!! Titles of tables go on TOP of the table while titles/captions of figures come BELOW the figure.

Connecting Results to Figures

From Kuroita, et al. "Structural mechanism for coordination of proofreading and polymerase activities in archaeal DNA polymerases." *JMB* 351, 2005, 291-298.

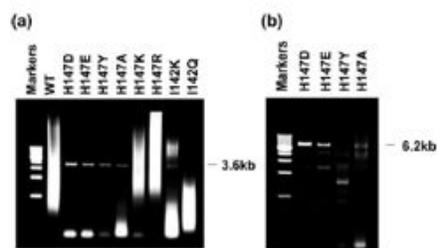


Figure 3. PCR with KOD polymerase mutants. (a) Agarose gel (1%) showing 3.6 kb PCR products. One unit of each mutant or WT enzyme was added to a mixture of 10 ng human genomic DNA and a primer pair designed to yield a 3.6 kb DNA fragment. (b) Long PCR with each mutant. One unit of each mutant was added to a mixture of 50 ng human genomic DNA and a primer pair designed to yield a 6.2 kb DNA fragment.

At first, a fragment of the human β -globin gene (3.6 kb) was amplified from different concentrations of human genomic DNA (final concentrations 2 ng/ μ l and 0.2 ng/ μ l) by each mutated enzyme. Under the high template DNA condition (2 ng/ μ l), each mutant showed a distinct band at the expected position upon gel analysis (data not shown). The change in template concentration from 2 ng/ μ l to 0.2 ng/ μ l greatly increased the frequency of failed reactions. Only four mutants (i.e. H147D, H147E, H147Y and H147A) resulted in successful amplification. Although H142K also showed a faint band, conspicuous unexpected bands were amplified at the same time. The other mutants generated only indistinct non-specific bands (Figure 3(a)). This experiment indicates that the 3'-5' exonuclease activity is not the only cause of PCR failure, because some mutants exhibiting similar Exo/Pol ratios (e.g. H147E and H142Q) produced different results. From these experiments, it is concluded that the negative charge or hydrophobicity of the amino acid at position 147 plays an important role for the sensitivity of PCR.

Next, the mutants that showed successful amplification in the above experiments (H147D, H147E, H147Y and H147A) were applied to "long PCR". A DNA fragment of the myosin heavy chain (6.2 kb) was amplified from human genomic DNA (final concentration, 1 ng/ μ l). As shown in Figure 3(b), H147D and H147E successfully amplified 6.2 kb products. The yield with H147D was higher than that with H147E. The target was not amplified by H147Y and H147A. PCR with the other mutants and the WT enzyme also ended in failure (data not shown). These results indicate that a negative charge at residue 147 of KOD DNA

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5.0 Discussion

With regards to the results obtained from flow cytometry, several unexpected results were observed. To begin with, all the negative controls had some cells that fell to the right of the diagonal line (greater FL1:FL2 ratio), suggesting that they expressed EGFP. This is likely due to the MES cells having background fluorescence or that there was contamination in the samples. However the most surprising result was the almost complete lack of homologous recombination in the 73+75SgrAI samples. This was surprising as we hypothesized that an increase in distance of a double strand break would decrease HR; however, we still believed that it would be greater than having no double strand breaks.

What features of this paragraph identify it as belong to the Discussion?

The purpose of the discussion section is to interpret and contextualize your data. You should begin by reiterating the purpose of your research and your major findings. Then you might do any or all of the following: connect your findings to other research (published or that of your peers); describe any ambiguities and sources of error in the data, and suggest future experiments to resolve uncertainties; explain where you expect your work may lead, and suggest specific experiments for extending your findings; describe any conceptual or technical limitations of the research. Finally, you should explain the significance of your findings to basic science and to engineering applications. Like the previous sections, the discussion should have a clear organization and narrative flow, whether or not you use sub-sections.

From http://openwetware.org/wiki/20.109%28S10%29:Guidelines_for_writing_up_your_research

Good MIT Resources

The Mayfield Guide On-Line

<http://www.mhhe.com/mayfieldpub/tsw/home.htm>

The MIT Writing and Communication Center

<http://web.mit.edu/writing/>

Writing and Speaking Resources on the 20.109 Wiki

Neal Lerner

[edit]

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Here are the slides (in PDF format) we showed in lab and class presentations:

September 17 and 18: [Overview of Scientific Writing and Rhetoric](#)

October 1 and 2: [Academic Integrity in Science and Engineering Communication](#)

Resources on Academic Integrity and Writing

[edit]

- [Academic Integrity at MIT](#) [@](#)
A rich site with examples and instruction on how to cite sources, paraphrase correctly, and avoid plagiarism.
- [From MIT's Office of Advising](#) [@](#)
Practical tips on avoiding plagiarism and keeping sane [@](#) MIT.
- [The OWL at Purdue on Plagiarism](#) [@](#)
- [Virginia Tech on Plagiarism](#) [@](#)
- [Norton on Plagiarism](#) [@](#)
- [Princeton on Plagiarism](#) [@](#)
- [Council of Writing Program Administrators on Plagiarism](#) [@](#)
- [Duke University Library on Plagiarism and Documentation](#) [@](#)

Writing Resources

[edit]

The following are helpful places to do further investigation into good writing.

- [The Mayfield Handbook](#) [@](#)
A rich resource for any scientific writer.
- [The MIT Writing and Communications Center](#) [@](#)
Located in 12-132, the Writing and Communications Center offers free one-to-one instruction on any aspect of writing.
- [The Science of Scientific Writing](#) [@](#)
A very useful article that analyzes the structure and style of scientific writing.
- [Writing Guidelines for Engineering and Science Students](#) [@](#)
A useful resource on scientific writing from Michael Alley, an engineering education professor at Penn State. Includes many examples and additional links.

Writing and Speaking Resources on the 20.109 Wiki

User:Atissa

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Atissa Banuazizi

[edit]

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Presentations

[edit]

All materials listed here are available as PDFs.

- [See my talk on Creating your 20.109 presentation.](#)
- [See my talk on Creating your research proposal presentation.](#)

Other resources

[edit]

- [Effective Presentations in Engineering and Science](#) [@](#) (Penn State)

MIT OpenCourseWare
<http://ocw.mit.edu>

20.109 Laboratory Fundamentals in Biological Engineering
Spring 2010

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